microEeze microbiological in-house test kits

CHROMAGAR S. AUREUS / CHROMAGAR S. AUREUS

DS049

hylabs

PRODUCT INFORMATION

Chromogenic medium for the isolation of Staphylococcus aureus.

PRINCIPLE

A color detection of S. aureus based on its enzymatic activity on chromogenic subtracts.

S. aureus gives mauve-pink colonies after 24 hours. Other microorganisms are inhibited or grow in blue, colorless or cream colonies.

The divided plate DD044, CHROMagar S. aureus and CHROMagar MRSA, with antibiotics addition at a final concentration of 4 mg cefoxitim litse/L, allows the isolation of methicillin resistant staphylococci.

INSTRUCTIONS FOR USE

Description

1. Plastic tube
2. Flexible plastic paddle with culture media on both sides, as detailed in the list below.
3. Screw plastic cap that may be taken apart from paddles.

Cat#	Description	Paddle Colour	Storage*
DS035	E. coli / Coliforms	Amber / Bordeaux	15-25°C
DS036	Salmonella / Salmonella *2	Pink, opaque	2-15°C
DS039	E. coli / E. coli	Amber	2-15°C
DS041	Aloa Agar Listeria / Aloa Agar Listeria *2	Yellow	2-15°C
DS049	S. Aureus / S. Aureus	Yellow	2-15°C
DS059	E. coli / S. Aureus	Amber / Yellow	2-15°C
DS068	CHROMagar STEC / Total Coliforms	Amber opaque / Bordeaux	2-15°C
DS063	CHROMagar E. coli 0157	Light Yellow	2-15°C

* Shipping Conditions: 2-25°C darkness

*2 These paddles are designed for surface contact. When using them by dipping method, a pre-enrichment stage has to be performed.

1. Remove paddle from tube by holding it by the cap. DO NOT TOUCH AGAR.
2. FOR LIQUIDS AND SOLIDS – (DIPPING METHOD) Dip paddle into sample completely covering the agar and remove immediately. If liquid is insufficient for dipping, pour liquid over the surface of both sides of the paddle. Liquid must be free flowing; syrupy samples or solids must be diluted up to 1:10 with

water so that the sample becomes free flowing. Drain excess liquid by wiping tip of the paddle on lip of the vessel containing the sample.

3. FOR SURFACES – (CONTACT SURFACE METHOD) Press the agar firmly on the surface to be tested for a few seconds, being careful not to smear the agar over the test area. Repeat the procedure using the second agar surface on a different site. Replace paddle in cap. Testing hands of employees: press fingers firmly on the agar surface.
4. Replace the paddle into the tube and close the cap loosely to allow for a free transfer of atmosphere. If the paddle is to be sent to a distant laboratory, close tightly and then upon arrival, open the cap loosely.
5. Write the relevant details on the identification label and stick on the tube.
6. Stand the tube in the incubator in an upright position with cap on top.
7. Read results, compare to chart and register as permanent record (taking into account any dilution performed in preparation of sample).
8. For Contact Surface Tests:
    a. Total Bacterial Count: Count the colonies on the agar surface and calculate according to the surface units.
    b. For Pathogens: Read the results as in the Table of Interpretation of Results (colony morphology) and proceed according to specifications for each product.

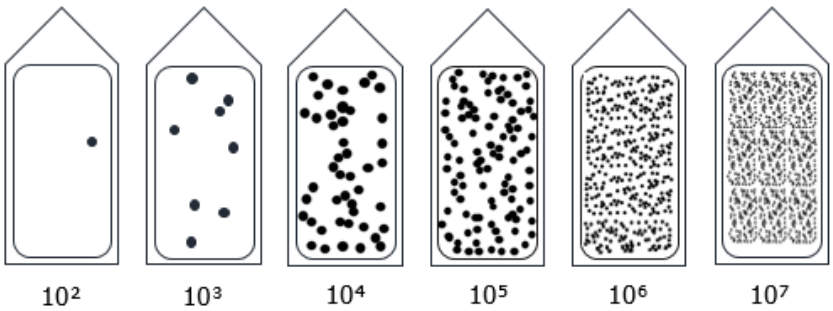
Incubation conditions and appearance of the colonies

Temperature*	**First Reading**	**Final Reading**	**Colony Color**
Coliforms – 30-35°C*	18 hours	24 hours	Only the pink colonies
E. Coli - 30-35°C*	18 hours	24 hours	Only the blue colonies
S. Aureus 35±2°C	----------	24 hours	Only the mauve colonies
Salmonella 35±2°C	----------	24 hours	Only the red colonies
Listeria 35±2°C	24 hours	48 hours	Only the green-turquoise colonies
CHROMagar STEC 35±2°C	----------	48 hours	Only the mauve colonies
CHROMagar E. Coli 0157 35±2°C	----------	24 hours	Only the mauve colonies

* Optimal temperatures

Interpretation of results

Dipping Test (c.f.u/ml)



Warning:

1. Used contaminated test material should be handled by standard decontamination methods such as: autoclaving, incineration or commercial bleach (hypochlorite) prior to disposal.
2. Any visibly contaminated or damaged goods should not be used and must be disposed of as in paragraph 1.

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